

Original Article

Plasma 17 β -estradiol and alkali-labile phosphoprotein levels in male and female Tench (*Tinca tinca*) in the Anzali and Amirkolayeh wetlands

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Abstract: Environmental pollutants are potentiate to disturb biological processes such as metabolism, growth and reproduction of aquatic organisms. These compounds are able to cause gonadal abnormalities, biased sex ratios and alteration in reproductive physiology in fish. The aim of this study was to examine plasma 17 β -estradiol (E2) and alkali-labile phosphoprotein (ALP) levels in male and female Tench (*Tinca tinca*) from a polluted (the Anzali Wetland) and a non-polluted environments (the Amirkolayeh Wetland). Samples were collected over the maturation season of Tench between May and June 2017. The results revealed significant difference in mean ALP and E2 between genders in the polluted environment. However, the mean plasma ALP concentrations in male Tench of the polluted environment (39.46 ± 1.02 $\mu\text{g/ml}$) was 45% of the average recorded in female (86.18 ± 2.25 $\mu\text{g/ml}$) and was two times higher than the amount measured in males in the non-polluted environment (18.68 ± 0.35 $\mu\text{g/ml}$). High concentrations of E2, were detected in the male samples from the Anzali Wetland. Mean plasma E2 concentrations for male in the Anzali Wetland was almost two times higher than male in the Amirkolayeh Wetland. The results indicate that the reproductive physiology of Tench was affected by contaminants found in the Anzali Wetland, a highly polluted area.

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Introduction

Fish reproductive physiology is controlled by the endocrine system, which is affected by environmental factors such as light, temperature, etc. This system includes various glands, synthesizing and secreting hormones, which in turn regulate fish reproduction (Pait and Nelson, 2003), control and initiate steroidogenesis, vitellogenesis and gametogenesis process mainly through the activation of hypothalamic–pituitary–gonadal (HPG) axis (Pankhurst and Munday, 2011). In aquatic systems, there are varieties of environmental pollutants causing disturbance in the function of this system (Ogundiran and Fawole, 2018) by simulating or blocking the steroid hormones activity (Mills and Chichester, 2005) and binding to estrogen or androgen receptors (Tapiero et al., 2002). These contaminants are called

endocrine disrupting chemicals (EDCs) and include groups of natural and synthetic compounds such as phytoestrogens (Burki et al., 2006), bisphenol-A (Qiu et al., 2015), PCBs (Walczak and Reichert, 2016) and PAHs (Vignet et al., 2016).

Releasing of large quantities of municipal, agriculture and industrial wastewaters into the aquatic ecosystems cause serious impacts on the physiology of aquatic organisms. The Anzali (Sakizadeh et al., 2012) and Amirkolayeh wetlands (Zare Khosh Eghbal and Sajadi Nasab, 2015) are located in the north of Iran and registered as two international wetlands based on 1975 Ramsar Convention. The main water source of the Amirkolayeh Wetland is underground water with no pollution from domestic and industrial wastes, and therefore, is considered as unpolluted and clean environment (Zare Khosh Eghbal and Sajadi

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Table 1. Total length and total weight (mean \pm SE) of Tench captured from the Anzali and Amirkolayeh wetlands (n=15 fish for each gender in each wetlands).

	Anzali Wetland		Amirkolayeh Wetland	
	Length (cm)	Weight (gr)	Length (cm)	Weight (gr)
Male	22.50 \pm 3.12	33.10 \pm 2.20	26.17 \pm 2.10	38.45 \pm 5.14
Female	25.13 \pm 6.41	40.87 \pm 4.89	28.32 \pm 4.52	43.19 \pm 7.26

Nasab, 2015). In contrast, concentrations of EDCs are considerably high in the Anzali Wetland (Mortazavi et al., 2012, 2013) due to releasing large amounts of untreated municipal and industrial wastewater into the wetland; as a result, fish could be exposed to the chemical compounds directly and indirectly in this wetland.

Recent investigations have suggested that exposure of wild fish to EDCs causes gonadal abnormalities and intersex (Scholz and Klüver, 2009), biased sex ratio (Larsson and Forlin, 2002) and induction of vitellogenin in males (Tyler et al., 2002). Vitellogenin is egg yolk protein precursor, which is synthesized in the liver of female fish (Jobling et al., 1996). Although there is vitellogenin gene in male, it is not expressed under normal conditions. However, when fish are exposed to EDCs, they may respond to these compounds by increasing the level of plasma vitellogenin. A considerable increase in the level of vitellogenin has been reported in male *Squalius cephalus* (Randak et al., 2009) and *Carassius auratus* (Li et al., 2009) inhabited in aquatic ecosystems contaminated by EDCs. Vitellogenin is dominant in plasma after the onset of vitellogenesis and the protein is heavily phosphorylated. This enables indirect quantification through measurement of alkali-labile protein bound phosphate (ALP) as alternative to the more expensive ELISA assay (Hallgren et al., 2009).

In this study, Tench (*Tinca tinca*) was selected as target species, being an indicator of water quality, used as a model species for studies of physiology (Martin et al., 1999), and lives in wetlands of Iran north, including the Anzali and Amirkolayeh wetlands. The present study started with hypothesis that reproductive physiology of Tench in the Anzali wetland could be affected by environmental pollution, and the Amirkolayeh wetland was considered as reference site.

Materials and Methods

Sixty male and female tench were collected during the reproduction season (between May and June 2017) by fyke nets (mesh size, 34-54 mm) at two sites: the Anzali Wetland (Pirbazar region), as a polluted environment and the Amirkolayeh Wetland as a non-polluted environment or reference site. Mean total length, sex, and number of collected samples are given in Table 1.

The fish were anesthetized in 150 mg/l clove powder solution and blood samples were immediately collected on site from the caudal vein using a heparinized 2.5 ml syringe. The collected blood from every fish was immediately placed in 2 ml plastic tubes, stored on ice, and transferred to the laboratory for further analyses. Blood samples were centrifuged at 1500 g, 4°C for 3 min (Scan Speed, 1730 R, Denmark). The separated plasma was stored at -80°C until analysis. Total length of fish were measured to the nearest millimeter, and weighed to the nearest gram. Gonads of each fish were observed macroscopically.

Plasma vitellogenine concentrations were measured indirectly by determination of plasma ALP (Negintaji et al., 2018; Matthiessen et al., 2018). In this method, vitellogenine-related phosphate released into plasma and was measured by spectrophotometer with the wavelength at 630 nm according to Hallgren et al. (2009). The levels of E2 were measured by Estradiol II kit (Biomereux, France) and with enzyme linked fluorescent assay (ELFA) method at 450 nm (Diver, 1987).

For calculating the effects of sex and location on measured parameters (ALP and E2), two-way analysis of variance (ANOVA) was applied. All data were tested for normality and homogeneity of variance using the Kolmogorov–Smirnov test and the Levene test, respectively. Statistical analyses were conducted

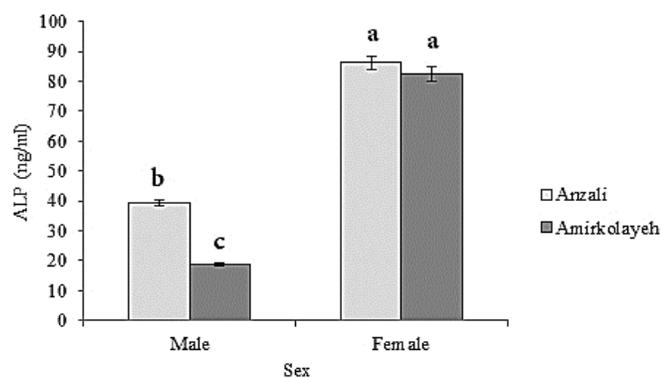


Figure 1. Concentrations of ALP for male and female Tench captured from the Anzali and Amirkolayeh wetlands (n = 15). Data are presented as the mean \pm standard error. Different letters above the bars show significant differences.

using SPSS (Version 16, Inc., Chicago, IL, USA). All statistics were performed with a critical α of 0.05.

Results

The males from the Anzali Wetland had significantly higher (~2 folds) ALP and E2 compared to those of the Amirkolayeh Wetland (Figs. 1, 2). There were no significant differences in the female's plasma ALP levels between the sampling sites (Fig. 1), but plasma E2 levels of the females from the Anzali Wetland were significantly higher than the females from the Amirkolayeh Wetland (Fig. 2). In both wetlands, females had significantly higher plasma ALP (Fig. 1) and E2 compared to the males (Fig. 2).

Discussions

There are increasing concerns for the presence of varieties of environmental pollutants with endocrine systems disrupting ability in aquatic organisms. EDC act by mimicking or interrupting hormone function, inducing vitellogenin synthesis in males, biased sex ratio and etc., which may affect natural reproductive and developmental processes (Mills and Chichester, 2005).

In the present study, the plasma ALP levels of the female fish from the Anzali Wetland were significantly different from males. Vitellogenin is a significant source of nutrition for developing embryos and larvae that synthesized in female fish liver in response to circulating E2 hormone (Monson et al.,

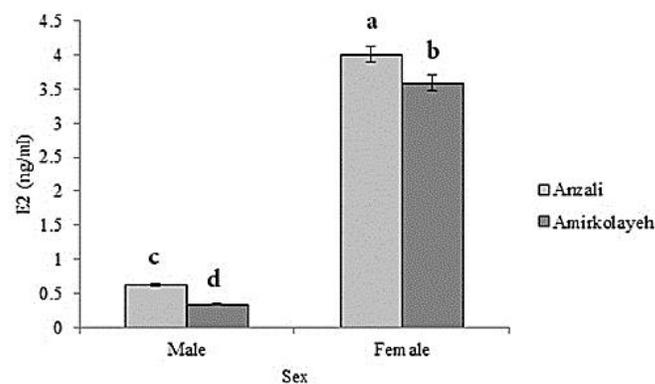


Figure 2. Concentrations of E2 for male and female of Tench captured from the Anzali and Amirkolayeh wetlands (n = 15). Data are presented as the mean \pm standard error. Different letters above the bars show significant differences. The bars show significant differences.

2017). However, during the reproductive cycle of Tench in the Anzali Wetland, synthesis vitellogenin was found in the male fish. There are vitellogenin genes in male fish and are not expressed under normal conditions, but might be induced when exposed to estrogen (Juin et al., 2017) or estrogen like compounds in the aquatic environment (Folmar et al., 2000). Field studies on wild *C. auratus* (Lu et al., 2010) and *Rhinichthys cataracte* (Jeffries et al., 2008) showed high concentrations of vitellogenin levels in the fish from EDCs-contaminated environment.

In this study, ALP levels of the males from the Anzali wetland were 45% of the females. Fossi et al. (2002) studied *Xiphias gladius* and *Thunnus thynnus* from the Mediterranean Sea, and found plasma vitellogenin concentrations of the males were 28% and 11% of the females, respectively. Plasma ALP levels in the males from the Anzali Wetland were higher than the males from the Amirkolayeh Wetland, which is similar to previous reports by Abbasi (2013) that suggested increased plasma ALP levels in the male pike (*Esox Lucius*) from the Anzali Wetland more than from Pike males captured from Amirkolayeh Wetland. Other researchers also found similar results over different species and in different regions (Hashimto et al., 2000; Folmar et al., 2001; Dick Vethaak et al., 2002).

Mortazavi et al. (2012, 2013) stated that the levels of EDCs (such as nonylphenol, octylphenol and bisphenol-A) at the Anzali Wetland were higher

compared to some other parts of the world such as rivers in China (Jin et al., 2010) and Japan (Isobe et al., 2001). When xenoestrogens present in water, they may change aromatase activity (the enzyme converts androgens to estrogens) in males by increasing plasma E2 concentrations (Solé et al., 2003) and induction of vitellogenin.

E2 is one of the most important sex steroids that are produced by ovarian follicles (Monson et al., 2017) and regulates sexual processing and reproductive processes (Martyniuk et al., 2006). Our results demonstrated that the males captured from the Anzali Wetland had a doubled E2 levels compared to the males from the Amirkolayeh Wetland, indicating that EDCs compounds at the Anzali Wetland interrupted reproduction system.

Conclusion

These results suggest that EDCs compounds in the Anzali Wetland might increase synthesis vitellogenin compared to the Amirkolayeh Wetland. The male Tench from the Anzali Wetland have increased plasma ALP and E2, suggesting disruption of reproduction system. However more studies will need to approve our results.

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References

Abbasi M. (2013). Vitellogenin index and sex ratio in Pike (*Esox lucius*) in the Anzali wetland. Master's thesis. University of Guilan, Faculty of natural resources. 57 p.

Burki R., Vermeirssen E.L., Körner O., Joris C., Burkhardt-Holm P., Segner H. (2006). Assessment of estrogenic exposure in brown trout (*Salmo trutta*) in a Swiss midland river: integrated analysis of passive samplers, wild and caged fish, and vitellogenin mRNA and protein. *Environmental Toxicology and Chemistry*, 25: 2077-2086.

Dick Vethaak A., Lahr J., Kuiper R.V., Grinwis G., Rankouhi T.R., Giesy J.P., Gerritsen A. (2002). Estrogenic effects in fish in The Netherlands: some preliminary results. *Toxicology*, 181: 147-150.

Diver M.J. (1987). Plasma estradiol concentration in neonates. *Clinical Chemistry*, 33: 1-17.

Folmar L.C., Denslow N.D., Kroll K., Orlando E.F., Enblom J., Marcino J., Metcalfe C., Guillette L.J. (2001). Altered serum sex steroids and vitellogenin induction in walleye (*Stizostedion vitreum*) collected near a metropolitan sewage treatment plant. *Archives of Environmental Contamination and Toxicology*, 40: 392-398.

Folmar L.C., Hemmer M., Hemmer R., Bowman C., Kroll K., Denslow N.D. (2000). Comparative estrogenicity of estradiol, ethynylestradiol and diethylstilbestrol in an in vivo, male sheepshead minnow (*Cyprinodon variegatus*), vitellogenin bioassay. *Aquatic Toxicology*, 49: 77-88.

Fossi M.C., Casini S., Marsili L., Neri G., Mori G., Ancora S., Moscatellic A., Ausilid A., Notarbartolo-di-Sciara G. (2002). Biomarkers for endocrine disruptors in three species of Mediterranean large pelagic fish. *Marine Environmental Research*, 54: 667- 671.

Hallgren P., Martensson L., Mathiasson L. (2009). Improved spectrophotometric vitellogenin determination via alkali-labile phosphate in fish plasma—a cost effective approach for assessment of endocrine disruption. *International Journal of Environmental and Analytical Chemistry*, 89: 1023-1042.

Hashimoto S., Bessho H., Hara A., Nakamura M., Iguchi T., Fujita K. (2000). Elevated serum vitellogenin levels and gonadal abnormalities in wild male flounder (*Pleuronectes yokohamae*) from Tokyo Bay, Japan. *Marine Environmental Research*, 49: 37-53.

Isobe T., Nishiyama H., Nakashima A., Takada H. (2001). Distribution and behavior of nonylphenol, octylphenol, and nonylphenol monoethoxylate in Tokyo metropolitan area: their association with aquatic particles and sedimentary distributions. *Environmental Science and Technology*, 35: 1041-1049.

Jeffries K.M., Nelson E.R., Jackson L.J., Habibi, H.R. (2008). Basin-wide impacts of compounds with estrogen-like activity on longnose dace (*Rhinichthys cataractae*) in two prairie rivers of Alberta, Canada. *Environmental Toxicology and Chemistry*, 27: 2042-2052.

Jin H., Chen J., Weng H., Li H., Zhang W., Xu J., Bai Y., Wang K. (2010). Variations in paleoproductivity and the environmental implications over the past six decades in the Changjiang estuary. *Acta Oceanologica*

- Sinica, 29: 38-45.
- Jobling S., Sumpter J.P., Sheahan D., Osborne J.A., Matthiessen, P. (1996). Inhibition of testicular growth in rainbow trout (*Oncorhynchus mykiss*) exposed to estrogenic alkylphenolic chemicals. *Environmental Toxicology and Chemistry*, 15: 194-202.
- Juin S. K., Mukhopadhyay B. C., Biswas S. R., Nath P. (2017). Conspecific vitellogenin induces the expression of vg gene in the Indian male walking catfish, *Clarias batrachus* (Linn.). *Aquaculture Reports*, 6: 61-67.
- Larsson D.J., Förlin L. (2002). Male-biased sex ratios of fish embryos near a pulp mill: Temporary recovery after a short-term shutdown. *Environmental Health Perspectives*, 110: 739-742.
- Li C.R., Lee S.H., Kim S.S., Kim A., Lee K.W., Lu M., Kim H.E., Kwak I.J., Lee Y.J., Kim D.K. (2009). Environmental estrogenic effects and gonadal development in wild goldfish (*Carassius auratus*). *Environmental Monitoring and Assessment*, 150: 397-404.
- Lu G.H., Song W.T., Wang C., Yan Z.H. (2010). Assessment of in vivo estrogenic response and the identification of environmental estrogens in the Yangtze River (Nanjing section). *Chemosphere*, 80: 982-990.
- Martin P., Rebollar P.G., Juan L.D.S., Illera J.C Alvarino J.M.R. (1999). Plasma estradiol-17 beta levels and gonadosomatic index in tench (*Tinca tinca* L.) Reared in natural and controlled conditions. *Journal of Physiology and Biochemistry*, 4: 309-314.
- Martyniuk C.J., Gallant N.S., Marlatt S.C., Woodhouse A.J., Trudeau V.L. (2006). Current perspectives on 17 β -estradiol action and nuclear estrogen receptors in teleost fish. In: M. Reinecke, G. Zaccane B. Kapoor, G. Enfield (Eds), *Fish endocrinology*. Science Publishers, New Hampshire. pp: 625-663.
- Masaru M., Bhandari R.K., Mikihiko H. (2003). The role estrogens play in sex differentiation and sex changes of fish. *Fish Physiology and Biochemistry*, 28: 113-117.
- Matthiessen P., Wheeler J. R., Weltje L. (2018). A review of the evidence for endocrine disrupting effects of current-use chemicals on wildlife populations. *Critical Reviews in Toxicology*, 48: 195-216.
- Mills L.J., Chichester C. (2005). Review of evidence: Are endocrine-disrupting chemicals in the aquatic environment impacting fish populations? *Science of the Total Environment*, 343: 1-34.
- Monson C., Forsgren K., Goetz G., Harding L., Swanson P., Young G. (2017). A teleost androgen promotes development of primary ovarian follicles in coho salmon and rapidly alters the ovarian transcriptome. *Biology of Reproduction*, 97: 731-745.
- Mortazavi S., Bakhtiari A.R., Sari A.E., Bahramifar N., Rahbarizadeh F. (2013). Occurrence of endocrine disruption chemicals (bisphenol A, 4-nonylphenol, and octylphenol) in muscle and liver of, *Cyprinus carpio* common, from Anzali Wetland, Iran. *Bulletin of Environmental Contamination and Toxicology*, 90: 578-584.
- Mortazavi S., Riyahi Bakhtiari A., Sari A.E., Bahramifar N., Rahbarizade F. (2012). Phenolic endocrine disrupting chemicals (EDCs) in Anzali Wetland, Iran: Elevated concentrations of 4-nonylphenol, octylphenol and bisphenol A. *Marine Pollution Bulletin*, 64: 1067-1073.
- Negintaji A., Safahieh A., Zolgharnein H., Matroodi S. (2018). Short-term induction of vitellogenesis in the immature male yellowfin seabream (*Acanthopagrus latus*) exposed to bisphenol A and 17 β -estradiol. *Toxicology and industrial health*, 34: 119-127.
- Ogundiran M.A., Fawole O.O. (2018). toxic effects of water pollution on two bio-indicators of aquatic resources of Asa River, Nigeria. *Journal of Fisheries Sciences*, 12: 20-27.
- Pait A.S., Nelson J.O. (2003). Vitellogenesis in male *Fundulus heteroclitus* (killifish) induced by selected estrogenic compounds. *Aquatic Toxicology*, 64: 331-342.
- Pankhurst N.W., Munday P.L. (2011). Effects of climate change on fish reproduction and early life history stages. *Marine and Freshwater Research*, 62: 1015-1026.
- Qiu W., Zhao Y., Yang M., Farajzadeh M., Pan C., Wayne N.L. (2015). Actions of bisphenol A and bisphenol S on the reproductive neuroendocrine system during early development in zebrafish. *Endocrinology*, 157: 636-647.
- Randak T., Zlabek V., Pulkrabova J., Kolarova J., Kroupova H., Siroka Z., Velisek J., Svobodova Z. Hajslova J. (2009). Effects of pollution on chub in the river Elbe, Czech Republic. *Ecotoxicology and Environmental Safety*, 72: 737-746.
- Sakizadeh M., Sari A.E., Abdoli A., Bahramifar N., Hashemi S.H. (2012). Determination of polychlorinated biphenyls and total mercury in two fish species (*Esox lucius* and *Carassius auratus*) in Anzali Wetland, Iran.

Environmental Monitoring and Assessment, 184: 3231-3237.

- Scholz S., Klüver N. (2009). Effects of endocrine disrupters on sexual, gonadal development in fish. *Sexual Development*, 3: 136-151.
- Solé M., Raldua D., Piferrer F., Barceló D., Porte C. (2003). Feminization of wild carp (*Cyprinus carpio*), in a polluted environment: plasma steroid hormones, gonadal morphology and xenobiotic metabolizing system. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 136: 145-156.
- Tapiero H., Nguyen G., Tew K.D. (2002). Estrogens and environmental estrogens. *Biomedicine and Pharmacotherapy*, 56: 36-44.
- Tyler C.R., Van Aerle R., Nilsen M.V., Blackwell R., Maddix S., Nilsen B.M., Berg K., Hutchinson T.H., Goksøyr A. (2002). Monoclonal antibody enzyme-linked immunosorbent assay to quantify vitellogenin for studies on environmental estrogens in the rainbow trout (*Oncorhynchus mykiss*). *Environmental Toxicology and Chemistry*, 21: 47-54.
- Vignet C., Larcher T., Davail B., Joassard L., Le Menach K., Guionnet T., Lyphout L., Ledevin M., Goubeau M., Budzinski H., Bégout M.L. (2016). Fish reproduction is disrupted upon lifelong exposure to environmental PAHs fractions revealing different modes of action. *Toxics*, 4: 1-26.
- Walczak M., Reichert M. (2016). Characteristics of selected bioaccumulative substances and their impact on fish health. *Journal of Veterinary Research*, 60: 473-480.
- Zare Khosh Eghbal M., Sajady Nasab M.S. (2015). Evaluation of pollution in Amir-kalayeh wetland sediment using geochemical analyses. *Journal of Environmental Geology*, 9: 1-10.

چکیده فارسی

سطوح ۱۷بتا-استرادیول و آلکالین-فسفو پروتئین پلازما در نر و ماده لای ماهی (*Tinca tinca*) در تالاب‌های انزلی و امیرکلايه

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چکیده:

آلاینده‌های زیست محیطی که به درون اکوسیستم‌های آبی وارد می‌شوند توانایی مختل کردن فرایندهای متابولیسم، رشد و نمو و تولیدمثل یک موجود آبی را داشته و می‌توانند موجب ناهنجاری‌های گنادی، انحراف نسبت‌های جنسی و تغییر فیزیولوژی تولیدمثل ماهی شوند. هدف از این مطالعه ارزیابی برخی پارامترهای بیوشیمیایی در پلاسمای خون لای ماهی نر و ماده از یک محیط آلوده (تالاب انزلی) و یک محیط غیرآلوده (تالاب امیرکلايه) می‌باشد. نمونه‌ها در طول فصل بلوغ لای ماهی بین ماه‌های اردیبهشت و خرداد ۱۳۹۶ جمع‌آوری شدند. فسفو پروتئین متصل به آلکالین (ALP) به عنوان یک شاخص غیرمستقیم از مقدار ویتلوژنین پلازما اندازه‌گیری شد. همچنین ۱۷-بتا استرادیول (E2) در نمونه‌های پلازما اندازه‌گیری شد. نتایج، اختلاف معنی‌داری در میانگین ALP و E2 بین جنس‌ها در محیط آلوده را نشان داد. با این حال غلظت‌های ALP در لای ماهی نر محیط آلوده ($39/46 \pm 1/02$ میکروگرم/میلی لیتر) ۴۵٪ میانگین ثبت شده در ماده‌ها ($86/18 \pm 2/25$ میکروگرم/میلی لیتر) و ۲ برابر مقدار اندازه‌گیری شده در لای ماهی‌های نر محیط غیر آلوده ($18/68 \pm 0/35$ میکروگرم/میلی لیتر) بود. غلظت‌های بالای E2 در نمونه‌های نر تالاب انزلی تعیین شد. میانگین غلظت‌های E2 پلازما در تالاب انزلی تقریباً دو برابر نرهای تالاب امیرکلايه بود. نتایج نشان داد که فیزیولوژی تولیدمثلی لای‌ماهی توسط آلاینده‌های تالاب انزلی، یک محیط به شدت آلوده، تحت تاثیر قرار گرفته است.

کلمات کلیدی: ALP، تالاب انزلی، هورمون استروئیدی، *Tinca tinca*.